

# Linked topological colloids in a nematic host

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Edited by John D. Weeks, University of Maryland, College Park, MD, and approved March 5, 2015 (received for review January 20, 2015)

3 4 114.1

-3

.4

-2-

-33-

. 2

-14.1 3 .2



r s r f l s . f l s r l s r d m



Ser nis =



l n l r nsr n (19) l r ,  
 t n r mn m n fl l s d s r ns nd d  
 m l - s d r l s fl l n k s d s fl  
 n r mn m fl r fl d n fl r ns ln rd fl l r s  
 (s m fl r l n k d r nd  
 l s). S r n r l n k n f d fl l s nd d l n k d  
 r n s r d s n d d n l n r n m n s m d  
 d fl ln ns n (F .4), s fl rd r f 150-70 fl r  
 s n l r fl n rd fl ln s (26, 30). D fl - n n l d l -  
 l d l n fl r ns, l k n s n n F .4, n  
 ll d l m n n r n s n r r (m s m-  
 m n l) n d nd n n m n fl d fl ln  
 ns n nd r ls l s fl r sd sr n dr rd s-  
 r ns n s r n s r n n  
 m n n r n s , ll d nd r s B n n m n  
 s n fl n s = 2 nd l d , s r l  
 d s n fl m fl s r l n s. B nd LC ll d s,  
 l n k n n l r n r ns n ll d l m n n s n,  
 fl r m l , n r ns r n fl m l r s rd l n  
 fl r s, n n n n fl r d s n n m l ll d l s fl  
 s s m l r r s nd fl n n l . n r nd, r  
 s s m l s l s d l s f d fl ln s m r d n  
 r m n l fl fl r m fl r s n rd ns fl m m l  
 m d l s fl r mn m m r ln fl l n k d nd k n d l l

r - rd r l n k s fl k n d l s fl s d ll d l r l s n  
 LCs, s r s n n l s m l s m l s fl sr d l -  
 l d l nd d r r sr r sd S l m n l n k s (F .8 nd  
 A , F .S4).  
 ns r r l fl l n k d r l m n n s ns d r l  
 n r s r s l r . n l k n n n n l ll d s,  
 m n - d n r ns n s m n l n k d m n n s  
 l n n s m m s r l , ns r nd  
 s l l n k n , r d fl r n m l m n n r l s. LC  
 l s - m d d n r ns n l n k d r n s fl  
 s m r l nd s r r nd n LC m r r d fl r n fl m  
 n d d fl s. Fr m l , n l n k n fl r n s s n n F .  
 2, fl r nd d ms r r n , s d r s l  
 n dr l r n r ns nd s fl - s s m l s m l r  
 s d d s l (19, 20), r d fl r n fl m l r m  
 nd m s l m l r r n m n s fl m n n r n s fl  
 l n k d r (F s. 2 nd 3). n l k n l n k d ll d l r n s  
 r r nd l r nd r nd ns, n  
 r n - l k nd n d fl s (19), r l n k d n r r s  
 n , n s r d nd n s n l r s. n l n k d l s  
 fl fl - n rd fl ln s (F .9) n , ll  
 l n n d fl s fl d r  $\pm 1$ , nd dd n l s fl  
 m ns n  $\pm 1$  n d fl r s n l s n r d d



r m n n n r n , n d n m n n s . n s ,  
 l n - k n r n s (33) h m n n s fl l-  
 l d l d n l s d m r . E r m n l s d fl  
 m l n s fl n n fl r l s n d d fl l s n r m s fl  
 l d d r n f l d d n l d t l s d l s l-  
 l r n m n s fl r n s m m r fl LC s s , . d s-  
 n s n n n l n d l n m s (34), s l l s  
 n d d r s fl s fl m r s s m s n r n l l  
 n r l n fl r n s (3, 7, 8, 19, 35-37). r n d d t l r  
 l r n s n l d s l n l s s n d s d fl s r r l s  
 d r m s , s l l s l l n f l n m n , k n r s s s , -  
 r l , s r f l n n r n l fl n k d l l d l m n n s , n d  
 r n l f l d s n r l m (35-37). L n k d m l m n n r-

laser light. The 3PEF-PM fluorescence intensity exhibits a strong well-defined dependence (25) on the orientation of linear polarization of the excitation beam relative to ( ). 3PEF-PM images, comprised of 3D stacks of optical slices, such as the ones shown in the SI Appendix, Figs. S12 and S13 and [Movies S3–S5](#), reveal orientations and relative positions of linked rings as well as the corresponding locations and configurations of topological defects accompanying them. Close analysis of 3PEF-PM stacks reveals dependence of 3D ( )-structures on boundary conditions and topology of colloids. Optical videomicroscopy and holographic laser tweezers (25) probe elastic interactions between the linked rings. Additionally, high-power beams of laser tweezers allow for locally

a 170- $\mu\text{m}$ -thick coverslip, spaced by 50- $\mu\text{m}$ -thick Mylar strips. As the beam focus was translated through the monomeric fluid, we always began polymerization at the substrate-fluid interface to effectively anchor the structure while it is being drawn. Arrays of particles were then detached from substrates by gentle sonication and dispersed into LCs. As-manufactured particles impose tangential boundary conditions for ( ), but some of them were treated with DMOAP for perpendicular ones (8).

**Optical Imaging and Laser Manipulation.** Director structures are studied using a combination of conventional polarizing optical microscopy and a 3D non-linear imaging technique dubbed “three-photon excitation fluorescence polarizing microscopy” (3PEF-PM) (25), which is based on fluorescence of LC molecules excited through three-photon absorption of femtosecond infrared